

Review

Role of Iron Metabolism in Anemia

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Abstract:

Anemia remains one of the most prevalent hematological disorders worldwide, affecting nearly two billion individuals. Central to its pathophysiology is iron metabolism, which underpins hemoglobin synthesis, mitochondrial energy production, and diverse enzymatic functions. Because unbound iron catalyzes reactive oxygen species formation, its regulation is tightly controlled through absorption, transport, storage, and recycling. Heparin, the master regulator of ferroportin, orchestrates systemic iron balance, and disturbances in this network give rise to distinct clinical phenotypes. Iron deficiency anemia commonly results from inadequate intake or chronic blood loss, whereas anemia of chronic disease reflects inflammatory upregulation of hepcidin. Genetic mutations in iron transport proteins further contribute to hereditary anemias. Recent advances emphasize the roles of hypoxia-inducible factors, erythropoietin, and immune signaling in adjusting iron availability during hypoxia and increased erythropoietic demand. Oxidative stress and ferroptosis exacerbate ineffective erythropoiesis and premature red cell destruction, linking iron imbalance to cellular injury. Macrophage-mediated recycling and hepatic storage normally buffer systemic fluctuations, but these mechanisms are impaired in chronic inflammation, thalassemia, and hemochromatosis. Clinically, precise diagnosis integrates conventional markers such as serum ferritin, transferrin saturation, and soluble transferrin receptor with emerging assays for hepcidin, which help distinguish iron deficiency anemia from anemia of chronic disease. Therapeutic innovation increasingly focuses on modulating hepcidin activity, targeting erythropoietin pathways, and developing safer iron formulations. Collectively, these insights highlight iron metabolism as a mechanistic cornerstone of anemia, bridging molecular regulation, oxidative stress, and biomarker development while guiding strategies for diagnostic accuracy and therapeutic progress.

Keywords: Anemia; Iron metabolism; Heparin; Erythropoietin; Hypoxia-inducible factors; Ferroptosis; Oxidative stress; Diagnostic biomarkers

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INTRODUCTION

Anemia is one of the most prevalent hematological disorders worldwide, affecting nearly two billion individuals and posing a major public health challenge across both developed and developing nations [1]. Anemia, marked by low hemoglobin or erythrocyte mass, impairs oxygen delivery; iron metabolism disturbances dominate its diverse etiological spectrum [2].

Iron supports hemoglobin, energy, DNA, and enzymes, yet excess causes oxidative stress. Regulation via DMT1, ferroportin, transferrin, ferritin, and hepcidin preserves systemic balance and cellular protection [3]. Together, these molecular players maintain iron equilibrium under physiological conditions. Thus, iron metabolism is not merely a nutritional concern but a mechanistic cornerstone in diverse anemic conditions [4]. Iron regulation remains complex; new studies highlight immune interplay, hypoxia-inducible factors,

and genetic diversity shaping individual susceptibility to iron imbalance [5].

Dysregulation of iron metabolism—particularly involving hepcidin activity, iron absorption, and mobilization from storage sites—contributes directly to the onset and progression of anemia by impairing erythropoiesis and limiting hemoglobin synthesis. Therefore, targeting key regulators of iron homeostasis may provide novel therapeutic strategies for anemia management

IRON METABOLISM : PHYSIOLOGICAL OVERVIEW

Physiological Overview of Iron Metabolism

Iron metabolism maintains balance by regulating absorption, distribution, storage, and recycling, ensuring essential functions while preventing harmful accumulation [6]. Iron absorption occurs mainly in the duodenum, involving heme and non-heme forms. Uptake, storage, and export via DMT1, ferritin, and ferroportin are tightly regulated by hepcidin to maintain systemic balance [7]. Most iron supports hemoglobin synthesis; surplus stored in ferritin and hemosiderin, while macrophages recycle iron from senescent red cells [8].

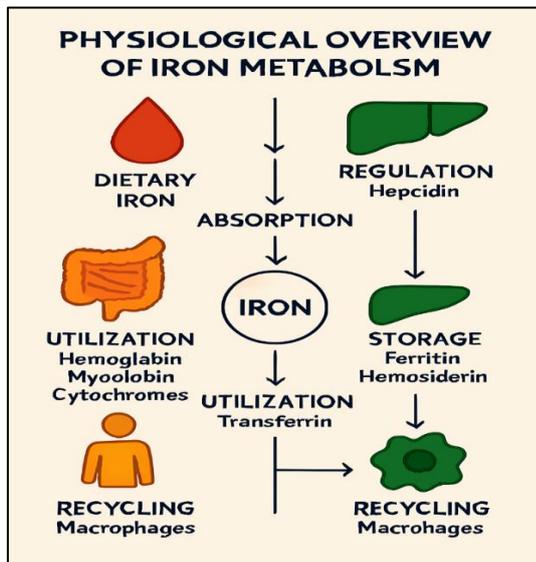


Fig 1: Physiological overview of iron metabolism

Absorption and Transport

Iron absorption in the duodenum involves heme and non-heme forms, differing in bioavailability and uptake, ensuring balance without toxicity [9]. Non-heme iron, ingested as ferric (Fe^{3+}), is reduced by Dcytb to ferrous (Fe^{2+}), absorbed via DMT1, with vitamin C enhancing and phytates, tannins, calcium inhibiting uptake [10]. Heme iron, highly bioavailable, enters enterocytes via HCP1 without reduction. Heme oxygenase liberates ferrous iron, enabling utilization or storage, explaining superior absorption from animal sources [11]. Iron is

oxidized to ferric (Fe^{3+}) by hephaestin and ceruloplasmin, then bound to transferrin, enabling safe transport, tissue delivery, and regulated uptake via transferrin receptors [12]. Iron absorption is tightly regulated since mammals lack excretion. Homeostasis relies on absorption and recycling; imbalance causes deficiency anemia or overload disorders, emphasizing coordinated intestinal, systemic, and hormonal control [13].

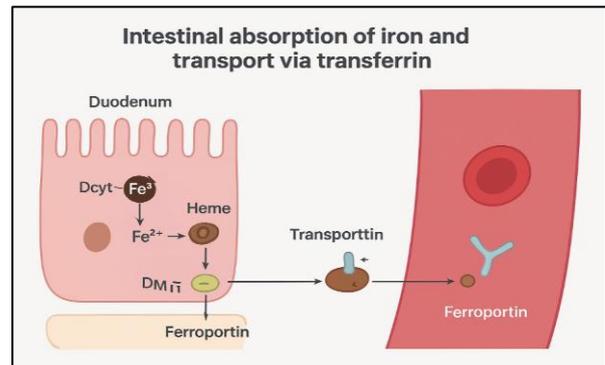


Fig 2: Intestinal absorption of iron and transport via transferrin

Cellular Regulation

Cellular iron uptake relies on transferrin receptor-mediated endocytosis. Acidic endosomes release ferric iron, reduced to ferrous by ferrireductases, then transported into the cytosol through divalent metal transporter [14]. Cytosolic iron is directed to mitochondria for heme and Fe-S cluster synthesis, supporting metabolism. Excess iron is sequestered in ferritin, preventing oxidative stress and maintaining cellular stability [15]. Ferritin and iron-related proteins are post-transcriptionally regulated by IRPs binding IREs. Low iron enhances uptake, while high iron promotes ferritin synthesis, ensuring cellular homeostasis [16]. Disrupted iron metabolism causes deficiency or overload, impairing mitochondria, inducing oxidative stress, and promoting ferroptosis linked to disease [17].

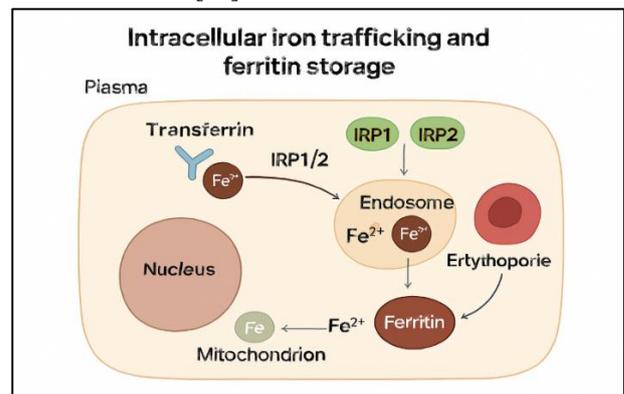


Fig 3: Intracellular iron trafficking and ferritin storage

Hormonal Control: Hepcidin

Iron supports hemoglobin, respiration, and enzymes; excess causes toxicity. Hepcidin, liver-derived hormone, regulates systemic iron balance to maintain homeostasis [18].

Regulation of Hepcidin Expression

Hepcidin synthesis is tightly regulated by multiple physiological signals:

- **Iron overload:** High plasma iron activates the BMP/SMAD signaling stimulating hepcidin transcription to prevent further absorption.
- **Inflammation:** Cytokines, particularly interleukin-6 (IL-6), upregulate hepcidin, contributing to anemia of chronic disease.
- **Erythropoietic demand:** Conditions such as anemia or hypoxia suppress hepcidin to ensure iron availability for hemoglobin synthesis [19].

Role of Erythroferrone

Erythroferrone, secreted by erythroblasts under erythropoietin stimulation, suppresses hepatic hepcidin, sustaining ferroportin activity, enhancing iron mobilization, and serving as biomarker and therapeutic target in iron-related disorders [20].

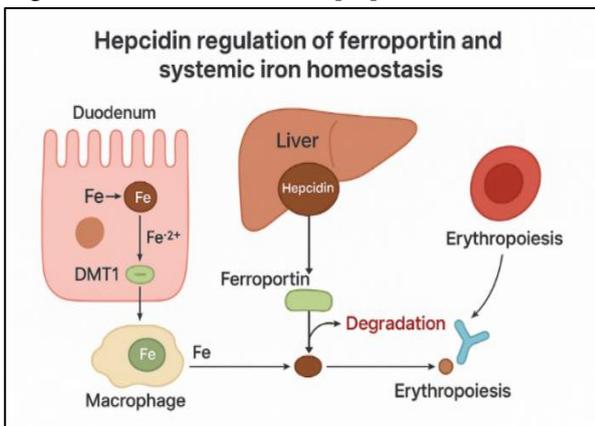


Fig 4: Hepcidin regulation of ferroportin and systemic iron homeostasis

Recycling Pathways

Macrophages recycle iron by engulfing aged erythrocytes, degrading hemoglobin, releasing heme, and exporting liberated iron via ferroportin, sustaining erythropoiesis and essential metabolic functions [21]. The liver stores iron in ferritin and hemosiderin, releasing it during deficiency. Along side macrophages, hepatic reserves regulate absorption, storage, and utilization, maintaining systemic iron balance [22].

Table 1: Types of Iron-Related to Anemia

Type of Anemia	Mechanism	Key Features
Iron Deficiency	Absolute lack of iron due to poor	Microcytic, hypochromic

Anemia (IDA)	intake, malabsorption, or chronic blood loss	RBCs; low ferritin; high TIBC [23]
Functional Iron Deficiency	Iron stores present but unavailable for erythropoiesis (often due to inflammation or hepcidin excess)	Normal/high ferritin but low transferrin saturation [24]
Anemia of Chronic Disease (ACD)	Cytokine-driven iron sequestration and reduced erythropoietin response	Normocytic or mildly microcytic; high ferritin; low serum iron [25]
Sideroblastic Anemia (Iron-loading anemia)	Defective incorporation of iron into heme	Ring sideroblasts in bone marrow; high serum iron [26]

Molecular Mechanism and Pathophysiology

a. Hepcidin regulation

Iron supports hemoglobin, respiration, and enzymes but excess is harmful. Hepcidin regulates ferroportin, coordinating absorption, storage, and mobilization to maintain systemic iron homeostasis [27].

- **BMP/SMAD Pathway: Primary Iron Sensing Mechanism** HJV, HFE, or TFR2 mutations lower hepcidin, elevate ferroportin, driving hereditary hemochromatosis with iron overload, cirrhosis, diabetes, cardiomyopathy, pigmentation [28].
- **JAK/STAT3 Pathway: Hepcidin in Inflammation** Interleukin-6 stimulates hepcidin transcription through JAK/STAT3 signaling. IL-6 activates JAKs, phosphorylating STAT3, which enters the nucleus, binds HAMP promoter, and enhances hepcidin gene expression [29]. Anemia of chronic disease arises from high hepcidin, causing macrophage iron sequestration, reduced absorption, functional deficiency with low serum iron yet elevated ferritin [30].
- **Suppression of Hepcidin: Hypoxia and Erythropoietic Demand** Erythroferrone from erythroblasts and hypoxia-inducible factors suppress hepcidin, sustaining ferroportin activity and iron mobilisation. This regulation supports erythropoiesis, while persistent suppression in

thalassemia promotes pathological iron overload [31].

- **Pathophysiological Consequences** Elevated hepcidin traps iron causing deficiency; reduced hepcidin overloads tissues, leading organ damage and disease [32].
- **Therapeutic Modulation of Hepcidin** Hepcidin agonists reduce iron absorption in overload, while antagonists alleviate inflammatory anemia. Therapeutic strategies also target BMP/SMAD and JAK/STAT3 pathways, offering promising avenues for regulating systemic iron homeostasis [33].

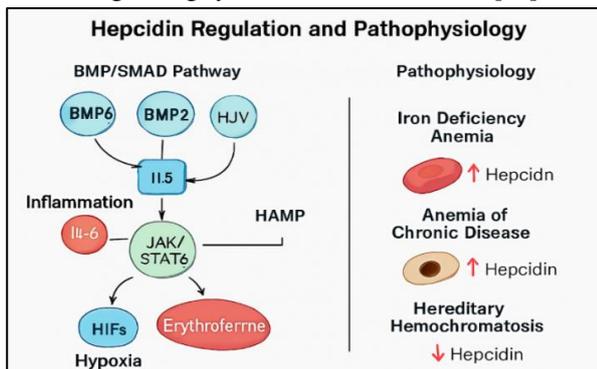


Figure 5: Hepcidine Regulation and Pathophysiology

b. Role of hypoxia-inducible factors (HIFs)

Hypoxia-inducible factors link oxygen sensing with iron metabolism; stabilized HIF-2 α enhances erythropoiesis by boosting erythropoietin, iron uptake, hepcidin suppression, and macrophage recycling, regulated via PHD-dependent oxygen-sensitive degradation [34]. A critical downstream target is erythropoietin (EPO). HIF-2 α induces EPO transcription in renal interstitial fibroblasts, stimulating erythropoiesis in the bone marrow. This mechanism compensates for hypoxic stress and is particularly relevant in anemia associated with chronic kidney disease, where impaired EPO production is a major contributor [35,36]. HIF-2 α enhances intestinal iron absorption by inducing DMT1 and DCYTB. DMT1 transports ferrous iron, while DCYTB reduces ferric, ensuring bioavailability and meeting erythropoietic demand through increased uptake [37]. Hepcidin limits iron by degrading ferroportin. HIFs suppress hepcidin via erythroferme and BMP modulation, enhancing plasma iron. In chronic disease anemia, cytokine-driven hepcidin elevation overrides suppression, restricting iron availability despite physiological demand [38,39]. Finally, HIFs regulate macrophage iron recycling by modulating polarization and erythrophagocytosis. Efficient recovery sustains erythropoiesis, while dysfunction under

hypoxia or inflammation impairs recycling, worsening anemia [40].

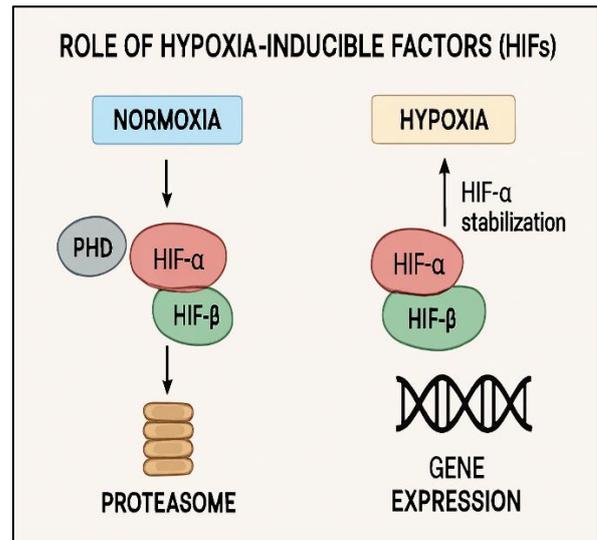


Figure 6: Role of HIF s

c. Erythropoiesis

- **EPO–JAK2–STAT5 Signaling** Erythropoietin drives erythropoiesis through EPOR–JAK2–STAT5, but anemia's inflammation and iron limits hinder output [41].
- **Iron Restriction and Hepcidin** Iron supports erythroblast heme synthesis. Hepcidin degrades ferroportin, restricting iron release. In chronic disease, cytokine-driven hepcidin elevation induces functional deficiency despite sufficient iron reserves [42].
- **Erythroferme (ERFE) Crosstalk** Erythroferme from EPO-stimulated erythroblasts suppresses hepcidin, enhancing iron for hemoglobin; chronic elevation in ineffective erythropoiesis causes overload despite anemia [43].
- **Hypoxia–HIF Pathway** Hypoxia stabilizes HIF-2 α , enhancing EPO transcription and iron transporter expression. Iron deficiency disrupts HIF-2 α via IRPs, diminishing EPO production and aggravating anemia through impaired oxygen–iron regulation [44].
- **Chromatin Remodeling in Erythroblasts** Terminal erythropoiesis requires chromatin condensation and enucleation; impaired heterochromatin remodeling disrupts maturation, causing anemia [45].
- **Macrophage Recycling** Macrophages recycle iron from aged erythrocytes, but inflammation-induced hepcidin blocks ferroportin, trapping iron and restricting erythropoiesis, causing anemia of chronic disease [46].

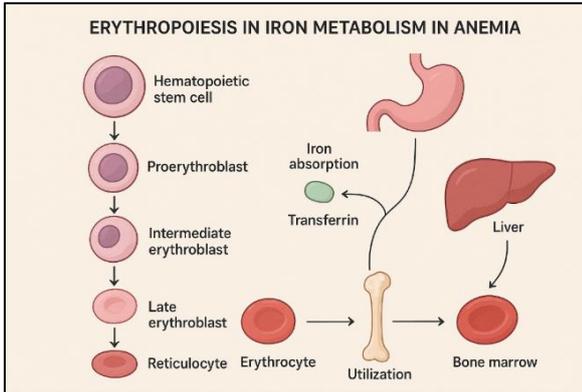


Fig 7: Erythropoiesis in Iron Metabolism in Anemia

d. Oxidative Stress

- **Iron as a Catalyst of ROS** Iron's redox cycling (Fe^{2+}/Fe^{3+}) drives Fenton/Haber–Weiss reactions, producing hydroxyl radicals that trigger oxidative damage—lipid peroxidation, protein oxidation, DNA injury—compromising erythrocyte viability [47].
- **Disruption of Iron Homeostasis** Iron homeostasis relies on ferritin, transferrin, and ferroportin. Oxidative stress disrupts them, releasing free iron, suppressing export, and elevating hepcidin, causing intracellular accumulation and impaired erythropoiesis [48].
- **Oxidative Damage to Erythrocytes** Reactive oxygen species damage erythrocyte membranes via lipid peroxidation and cytoskeletal injury, inducing
 - 1) **Diagnostic markers**

eryptosis with shrinkage, blebbing, phosphatidylserine exposure, ultimately reducing lifespan and promoting anemia [49].

- **Mitochondrial Dysfunction in Erythropoiesis** Erythroblast mitochondria synthesize heme and iron–sulfur clusters; oxidative stress disrupts enzymes, impairing hemoglobin, leading to thalassemia and sideroblastic anemia [50].
- **Ferroptosis and Iron-Driven Cell Death** Ferroptosis via iron-driven lipid peroxidation impairs erythroid precursors, aggravating ineffective erythropoiesis and anemia progression [51].

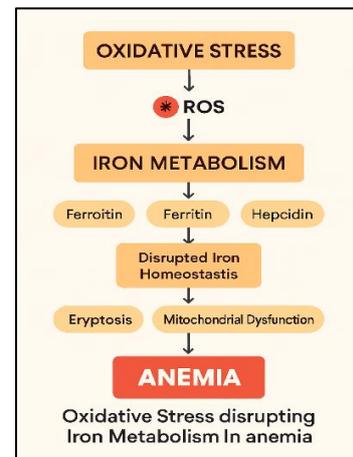


Fig 8: Oxidative Stress disrupting Iron Metabolism In Anemia

Table 2: Diagnostic markers and their Role

Marker	Physiological Role	Diagnostic Utility	Limitations
Serum Ferritin	Storage protein for iron; reflects body iron reserves	Low ferritin (<30 ng/mL) is highly specific for IDA	Elevated in inflammation, liver disease, and malignancy (acute-phase reactant) [52]
Transferrin Saturation (TSAT)	Ratio of serum iron to total iron-binding capacity	TSAT < 20% suggests iron deficiency; useful for iron overload	Fluctuates with diurnal variation, inflammation, malnutrition [53]
Soluble Transferrin Receptor (sTfR)	Circulating fragment of transferrin receptor; reflects erythropoietic iron demand	Elevated in IDA; unaffected by inflammation; sTfR/log ferritin index distinguishes IDA vs ACD	Limited availability, higher cost, assay variability [54]

• **Emerging biomarkers:**

Hepcidin assays Hepcidin controls iron via ferroportin degradation; high restricts absorption, low enhances uptake, aiding anemia management [55].

Types of Hepcidin Assays

1. Mass Spectrometry (MS) Assays
2. Immunoassays (ELISA, RIA)
3. Novel Immunological Assays

2) Therapeutic strategies: oral/parenteral iron, erythropoiesis-stimulating agents, hepcidin modulators.

○ **Oral Iron Therapy**

Recent Oral Formulations

- Ferric maltol: improved bioavailability and tolerability.

- Sucrosomial iron: encapsulated formulation with better GI tolerance, useful in patients intolerant to ferrous salts [56].

- **Parenteral (Intravenous) Iron Therapy**

Formulations

- Iron sucrose
- Ferric carboxymaltose (FCM) – allows high-dose single infusions.
- Ferumoxytol – rapid infusion, lower hypersensitivity risk.
- Iron dextran – older, associated with higher risk of allergic reactions [57].

- **Recent Drugs in Anemia Therapy**

Beyond traditional iron supplementation, new pharmacological agents have emerged:

Hypoxia-Inducible Factor Prolyl Hydroxylase Inhibitors (HIF-PHIs)

- Daprodustat (Jesduvrog) – FDA-approved (2023) for CKD-related anemia.
- Roxadustat – approved in Asia and Europe; lowers hepcidin, improves iron mobilization.
- Vadadustat – approved in Japan/EU; oral alternative to ESAs.
- Mechanism: Stabilize hypoxia-inducible factors → increase endogenous erythropoietin, enhance iron metabolism [58].

Novel Iron Formulations

- Ferric carboxymaltose (FCM) – high-dose IV iron, fewer infusions.
- Ferumoxytol – rapid infusion, safer profile.
- Sucrosomial iron – oral, better GI tolerance [59].

Hepcidin Modulators

- **LY2787106 (anti-hepcidin antibody)** – restores iron availability in anemia of inflammation.
- **PTG-300 (rusfertide)** – hepcidin mimetic, under trial for iron overload but with anemia potential.
- **Spiegelmers (NOX-H94)** – oligonucleotide-based hepcidin antagonists [60].

Recent Advance Treatments

Nanomedicine in iron delivery Nanomedicine in iron therapy employs nanoparticles to enhance absorption, distribution, and efficacy, addressing limitations of conventional formulations like poor bioavailability, gastrointestinal discomfort, and oxidative stress, thereby improving therapeutic outcomes in anemia management[61].

Gene therapy for iron metabolism disorders

Gene therapy revolutionises iron disorder treatment by correcting genetic defects in hepcidin–ferroportin, transferrin signaling, and hemoglobin synthesis, offering lasting cures beyond conventional symptom management through precise gene editing or replacement [62]. Iron metabolism disorders stem from genetic mutations; gene therapy provides curative potential by correcting molecular defects beyond conventional treatments [63].

Methods of Gene Therapy in Iron Metabolism Disorders

- 1) **Gene Addition (Replacement Therapy)** Functional copies of defective genes are introduced using viral vectors such as lentiviruses or adeno-associated viruses. Example: Adding normal *HAMP* or *TFR2* genes to restore hepcidin regulation and iron sensing [64].
- 2) **Gene Editing (Correction Therapy)** CRISPR-Cas9, TALENs, and zinc finger nucleases enable precise correction of mutations. In β -thalassemia, CRISPR-mediated editing of *BCL11A* enhances fetal hemoglobin, indirectly balancing iron metabolism [65].
- 3) **Gene Silencing (Downregulation Therapy)** RNA interference and antisense oligonucleotides are used to silence pathogenic gene expression. Silencing *HAMP* in conditions of excessive hepcidin may restore iron availability [66].
- 4) **Ex Vivo Stem Cell Editing** Hematopoietic stem cells are harvested, genetically corrected outside the body, and reinfused into patients. This approach is being tested in clinical trials for β -thalassemia and sickle cell disease [67].
- 5) **In Vivo Delivery Systems** Nanoparticle-based CRISPR delivery and improved viral vectors are being developed to directly target hepatocytes and bone marrow cells. This method aims to bypass the need for stem cell transplantation [68].

Novel hepcidin antagonists and agonists

Hepcidin modulators adjust iron balance through ferroportin and BMP pathways, offering therapeutic potential in diverse anemias [69].

Hepcidin Antagonists Hepcidin antagonists block hepcidin-ferroportin interaction, enhancing iron availability for erythropoiesis, treating anemia of chronic disease, IRIDA, and β -thalassemia [70].

- **Monoclonal Antibodies Against Hepcidin** Neutralizing antibodies block hepcidin-ferroportin binding, restoring iron and hemoglobin in inflammatory anemia models; ongoing trials assess efficacy in chronic kidney disease and cancer-related anemia patients [71].
- **Ferroportin Stabilizers** Small molecules inhibit hepcidin-ferroportin binding, stabilizing ferroportin, sustaining iron export, and offering therapeutic potential for IRIDA and inflammatory anemia [72].
- **BMP/SMAD Pathway Inhibitors** BMP6-SMAD signaling drives hepcidin transcription; inhibitors of BMP receptors or SMAD reduce synthesis, enhancing iron absorption. Dorsomorphin derivatives and anti-BMP6 antibodies are currently under investigation [73].
- **TMPRSS6 Agonists** TMPRSS6 cleaves hemojuvelin, reducing hepcidin transcription; pharmacological activation enhances iron absorption, offering therapeutic relevance for IRIDA linked to TMPRSS6 mutations [74].
- **RNA-Based Therapies** Antisense oligonucleotides and siRNAs suppress HAMP mRNA, lowering hepcidin, enhancing iron availability, and showing promise for treating anemia of inflammation [75].

Hepcidin Agonists Hepcidin agonists aim to increase hepcidin activity or mimic its function, thereby reducing iron overload. They are particularly useful for hereditary hemochromatosis, transfusional iron overload, and β -thalassemia with ineffective erythropoiesis [76].

- **Synthetic Hepcidin Peptides** Engineered analogs of hepcidin can directly bind ferroportin, mimicking natural hormone activity. These peptides reduce serum iron and tissue iron deposition in animal models of hemochromatosis [77].
- **Mini-Hepcidins** Mini hepcidins, stable peptide derivatives with improved bioavailability, ease administration, reduce iron overload in preclinical models, and are currently undergoing clinical trials for human safety and efficacy [78].
- **Gene Therapy Approaches** Viral vector delivery of the *HAMP* gene or CRISPR-based upregulation of hepcidin expression in hepatocytes offers durable

correction. This strategy could provide long-term control of iron overload in hereditary hemochromatosis [79].

- **Small-Molecule Inducers** Compounds that activate BMP6/SMAD signaling increase endogenous hepcidin production. These agents are being explored as oral therapies for iron overload disorders [80].
- **Hepcidin Mimetics** Engineered antibodies or fusion proteins that replicate hepcidin's ferroportin-binding effect are being developed as long-acting agonists. These mimetics may provide sustained iron regulation with fewer dosing requirements [81].
- **Systems Biology Approaches** Systems biology integrates computational modeling, high-throughput omics technologies, and network analysis to understand complex biological processes at a holistic level [82].

Conclusion

Anemia continues to represent a global health challenge, with iron metabolism at the core of its pathophysiology. Iron is indispensable for hemoglobin synthesis, cellular respiration, and enzymatic activity, yet its dual nature—as both essential and potentially toxic—necessitates precise regulation. The interplay of absorption, transport, storage, and recycling, coordinated by hepcidin and ferroportin, ensures systemic balance under normal conditions. Disruption of this network manifests in diverse clinical forms, ranging from iron deficiency anemia to anemia of chronic disease and genetic iron-loading disorders. Recent advances have expanded our understanding of iron biology, highlighting the roles of hypoxia-inducible factors, erythroferrone, and immune signaling in modulating iron availability. Oxidative stress and ferroptosis further illustrate how iron imbalance contributes to ineffective erythropoiesis and premature erythrocyte destruction. These mechanistic insights not only deepen our comprehension of anemia but also reveal novel therapeutic targets. Hepcidin modulation, erythroferrone regulation, and innovative iron formulations exemplify translational strategies aimed at restoring iron equilibrium and improving patient outcomes. From a clinical perspective, integrating conventional markers such as ferritin, transferrin saturation, and soluble transferrin receptor with emerging

hepcidin assays enhances diagnostic precision, particularly in distinguishing iron deficiency anemia from anemia of chronic disease. This diagnostic refinement is critical for guiding appropriate therapy and avoiding mismanagement. In summary, iron metabolism is a mechanistic cornerstone in anemia research and clinical practice. Continued exploration of its regulatory pathways and pathological consequences will not only advance scientific knowledge but also foster innovative interventions to alleviate the global burden of anemia.

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REFERENCES:

- World Health Organization. Guidance on implementing patient blood management to improve global blood health status. World Health Organization; 2025 Mar 13.
- Kolarš B, Mijatović Jovin V, Živanović N, Minaković I, Gvozdrenović N, DickovKokeza I, Lesjak M. Iron deficiency and iron deficiency anemia: A comprehensive overview of established and emerging concepts. *Pharmaceuticals*. 2025 Jul 25;18(8):1104.
- Pagani A, Nai A, Silvestri L, Camaschella C. Heparin and anemia: a tight relationship. *Frontiers in physiology*. 2019 Oct 9;10:1294.
- Coates TD, Cazzola M. Introduction to a review series on iron metabolism and its disorders. *Blood, The Journal of the American Society of Hematology*. 2019 Jan 3;133(1):1-2.
- Malik ZI, Ghafoor MU, Shah SH, Abid J, Farooq U, Ahmad AM. Unlocking iron: nutritional origins, metabolic pathways, and systemic significance. *Frontiers in Nutrition*. 2025 Aug 13;12:1637316.
- Vargas-Almendra A, Ruiz-Medrano R, Núñez-Muñoz LA, Ramírez-Pool JA, Calderón-Pérez B, Xoconostle-Cázares B. Advances in soybean genetic improvement. *Plants*. 2024 Oct 31;13(21):3073.
- Zhong M, Wang Y, Min J, Wang F. Iron metabolism and ferroptosis in human health and disease. *BMC biology*. 2025 Aug 22;23(1):263.
- Horváth A, Tamási K, Pap R, Jánosa G, Pandur E. Iron, the Essential Micronutrient: A Comprehensive Review of Regulatory Pathways of Iron Metabolism. *Nutrients*. 2025 Dec 28;18(1):109.
- Chen X, Yu C, Kang R, Tang D. Iron metabolism in ferroptosis. *Frontiers in cell and developmental biology*. 2020 Oct 7;8:590226.
- Ru Q, Li Y, Chen L, Wu Y, Min J, Wang F. Iron homeostasis and ferroptosis in human diseases: mechanisms and therapeutic prospects. *Signal transduction and targeted therapy*. 2024 Oct 14;9(1):271.
- Ganz T, Nemeth E. Iron homeostasis in host defence and inflammation. *Nature Reviews Immunology*. 2015 Aug;15(8):500-10.
- Anderson GJ, Frazer DM. Current understanding of iron homeostasis. *The American journal of clinical nutrition*. 2017 Dec 1;106:1559S-66S.
- Li C, Zhou L, Yin X. Pathophysiological aspects of transferrin-A potential nano-based drug delivery signaling molecule in therapeutic target for varied diseases. *Frontiers in Pharmacology*. 2024 Mar 4;15:1342181.
- Guo Q, Qian C, Wang X, Qian ZM. Transferrin receptors. *Experimental & Molecular Medicine*. 2025 Apr 22:1-9.
- Vogt AC, Arsiwala T, Mohsen M, Vogel M, Manolova V, Bachmann MF. On iron metabolism and its regulation. *International journal of molecular sciences*. 2021 Apr 27;22(9):4591.
- Sun S, Shen J, Jiang J, Wang F, Min J. Targeting ferroptosis opens new avenues for the development of novel therapeutics. *Signal transduction and targeted therapy*. 2023 Sep 21;8(1):372.
- Babitt JL. Erythroferrone in iron regulation and beyond. *Blood, The Journal of the American Society of Hematology*. 2022 Jan 20;139(3):319-21.
- Babker AM. The associations between erythroferrone and hepcidin during iron deficiency anemia, iron overload and pregnancy. *International Journal of Health Sciences*. 2022(1):6171-7.
- Camaschella C. Iron deficiency and iron overload: from pathophysiology to clinical management. *Haematologica*. 2021;106(8):2208-2219.
- Ganz T, Nemeth E. Heparin and iron homeostasis in health and disease. *Blood*. 2021;138(20):1616-1624.

- 22 Nemeth E, Ganz T. Hepcidin and iron in health and disease. *Annual review of medicine*. 2023 Jan 27;74(1):261-77.
- 23 El-Gamal RA, Abdel-Messih IY, Habashy DM, Zaiema SE, Pessar SA. Erythroferrone, the new iron regulator: evaluation of its levels in Egyptian patients with beta thalassemia. *Annals of Hematology*. 2020 Jan;99(1):31-9.
- 24 Nemeth E, Ganz T. Introduction to a review series on iron metabolism and its disorders. *Blood*. 2019;133(1):1-6.
- 25 Rolić T, Yazdani M, Mandić S, Distanto S. Iron metabolism, calcium, magnesium and trace elements: a review. *Biological Trace Element Research*. 2025 Apr;203(4):2216-25.
- 26 Saad HKM, et al. Activation of STAT and SMAD Signaling Induces Hepcidin Re-Expression as a Therapeutic Target for β -Thalassemia Patients. *Biomedicine*. 2022;10(1):189.
- 27 Charlebois E, Pantopoulos K. Iron overload inhibits BMP/SMAD and IL-6/STAT3 signaling to hepcidin in cultured hepatocytes. *PLoS One*. 2021;16(6):e0253475.
- 28 Girelli D, Nemeth E, Swinkels DW. Hepcidin in the diagnosis of iron disorders. *Blood*. 2021;138(22):2377-2389.
- 29 Jayakumar D, Narasimhan KK, Periandavan K. Triad role of hepcidin, ferroportin, and Nrf in cardiac iron metabolism: From health to disease. *Journal of Trace Elements in Medicine and Biology*. 2022 Jan 1;69:126882..
- 30 Camaschella C. Iron deficiency and iron overload: from pathophysiology to clinical management. *Haematologica*. 2021;106(8):2208-2219.
- 31 Coffey R, Babitt JL. Erythroferrone in iron regulation and beyond. *Blood*. 2022;139(3):319-321.
- 32 Arezes J, Nemeth E. Hepcidin and iron disorders: new insights into pathophysiology and treatment. *Nat Rev Gastroenterol Hepatol*. 2020;17(7):407-418.
- 33 Nai A, et al. Hepcidin regulation and clinical implications in hematology. *Curr Opin Hematol*. 2020;27(4):183-190.
- 34 Semenza GL. Hypoxia-inducible factors in physiology and medicine. *Blood*. 2022;139(16):2441-2449.
- 35 Haase VH. Regulation of erythropoiesis by hypoxia-inducible factors. *Blood Rev*. 2021;50:100853.
- 36 Chen N, Hao C, Liu BC, et al. Roxadustat treatment for anemia in chronic kidney disease. *Kidney Int*. 2021;99(6):1436-1447.
- 37 Gupta N, Wish JB. Hypoxia-inducible factor stabilizers: emerging therapy for anemia. *Clin J Am Soc Nephrol*. 2020;15(9):1438-1448.
- 38 Locatelli F, Fishbane S, Block GA, et al. Targeting HIF pathway in renal anemia: clinical trials update. *Nephrol Dial Transplant*. 2021;36(12):2103-2111.
- 39 Zhang J, Wang Y, Xu J, et al. HIF pathway modulation in anemia of chronic disease. *Int J Mol Sci*. 2022;23(14):7654.
- 40 Wang GL, Jiang BH. HIF-2 α and iron metabolism in anemia. *Front Med*. 2022;16(3):321-330.
- 41 Vives-Corrons JL, Krishnevskaya E. Hereditary Spherocytosis: Linking Ion Transport Defects to Osmotic Gradient Ektacytometry Profiles—A Review. *International Journal of Molecular Sciences*. 2026 Jan 10;27(2):721.
- 42 Camaschella C, Pagani A, Silvestri L, Nai A. The mutual crosstalk between iron and erythropoiesis. *Int J Hematol*. 2022;116:695-707.
- 43 Li D, Wu F, Zhou S, Huang XJ, Lee HY. Heterochromatin rewiring and domain disruption-mediated chromatin compaction during erythropoiesis. *Nat Struct Mol Biol*. 2023;30:345-357.
- 44 Ginzburg Y, An X, Rivella S, Goldfarb A. Normal and dysregulated crosstalk between iron metabolism and erythropoiesis. *eLife*. 2023;12:e90189.
- 45 Pasricha SR, Tye-Din J, Muckenthaler MU. Iron deficiency and anemia: molecular insights and clinical implications. *Lancet Haematol*. 2023;10:e98-e110.
- 46 Galaris D, Barbouti A, Pantopoulos K. Iron homeostasis and oxidative stress: An intimate relationship. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*. 2019 Dec 1;1866(12):118535.
- 47 Iuchi Y. Anemia caused by oxidative stress. *Anemia*. 2012 Feb 29.
- 48 Zhong A, Yan X. Erythropoiesis in teleost fishes: the fantastic biological process. *Reviews in Aquaculture*. 2025 Jan;17(1):e12960.
- 49 Wang Q, Gao R, Zhu K, Qiu H, Huang J, Zhang X. The Potential Role of Iron Homeostasis and Ferroptosis in Exercise Nutrition and Health. *Nutrients*. 2026 Jan 1;18(1):139.
- 50 Kesharwani P, Dash D, Koiri RK. Deciphering the role of hepcidin in iron metabolism and anemia

- management. *Journal of Trace Elements in Medicine and Biology*. 2025 Jan 8:127591.
- 51 Rusch JA, van der Westhuizen DJ, Gill RS, Louw VJ. Diagnosing iron deficiency: Controversies and novel metrics. *Best Pract Res Clin Anaesthesiol*. 2025;37(4):451-467.
 - 52 Skikne BS, Punnonen K, Caldron PH, Bennett MT, Rehu M, Gasior GH. Improved differential diagnosis of anemia of chronic disease and iron deficiency anemia: a prospective multicenter evaluation of soluble transferrin receptor and the sTfR/log ferritin index. *American journal of hematology*. 2011 Nov;86(11):923-7.
 - 53 Nadeem S, Shah S, Iqbal T, et al. Serum transferrin receptor, serum ferritin and serum transferrin receptor-ferritin index in adults with iron deficiency anemia. *J Ayub Med Coll Abbottabad*. 2011;23(3):77-81.
 - 54 Łęcka M, Słomka A, Albrecht K, Romiszewski M, Styczyński J. Iron dysregulation signature in pediatric leukemia: in-depth biomarkers of iron metabolism involving matrilysin-2 and neogenin-1. *Cancers*. 2025 Jul 29;17(15):2495.
 - 55 Fathi ZH, Mohammad JA, Younus ZM, Mahmood SM. Heparin as a Potential Biomarker for the Diagnosis of Anemia. *Turkish journal of pharmaceutical sciences*. 2022 Oct 31;19(5):603.
 - 56 Rana S, Prabhakar N. Iron disorders and hepcidin. *Clinica chimica acta*. 2021 Dec 1;523:454-68.
 - 57 Rana S, Prabhakar N. Iron disorders and hepcidin. *Clinica chimica acta*. 2021 Dec 1;523:454-68.
 - 58 Myshko D. FDA Approves Novel Therapy for CKD-Related Anemia. *Formulary Watch*. 2023 Feb 2:NA-.
 - 59 Auerbach M, Macdougall IC. Safety of intravenous iron formulations: facts and folklore. *Blood Transfus*. 2014;12(3):296-300.
 - 60 Ghazi R, Ibrahim TK, Nasir JA, Gai S, Ali G, Boukhris I, Rehman Z. Iron oxide based magnetic nanoparticles for hyperthermia, MRI and drug delivery applications: a review. *RSC advances*. 2025;15(15):11587-616.
 - 61 Omoruyi IC, Omoruyi JI, Aghedo ON, Archibong UD, Ifijen IH. Application of magnetic iron oxide nanostructures in drug delivery: a compact review. In *TMS Annual Meeting & Exhibition 2023 Feb 7* (pp. 229-242). Cham: Springer Nature Switzerland.
 - 62 Li Y, Barmin RA, Zhang R, Kiessling F, Lammers T, Pallares RM. Clinical translation and landscape of superparamagnetic iron oxide nanoparticles. *Advanced Drug Delivery Reviews*. 2025 Dec 6:115756.
 - 63 Losito DW, Souza NI, Martins TS, Britos TN, Schumacher ML, Haddad PS. A review of superparamagnetic nanoparticles applications and regulatory aspects in medicine and environmental areas. *Journal of Materials Science*. 2024 Sep;59(34):16038-68.
 - 64 Mansour GK, Hajjar AW, Sajid MR. Therapeutic targeting of the hepcidin-ferroportin axis and erythropoietic modulators: a narrative review. *Frontiers in Medicine*. 2025 Dec 9;12:1726337.
 - 65 Li L, Mandal PK. Recent advancements in gene therapy for sickle cell disease and β -thalassemia. *Frontiers in Hematology*. 2024 Sep 27;3:1468952.
 - 66 Alayoubi AM, Khawaji ZY, Mohammed MA, Mercier FE. CRISPR-Cas9 system: a novel and promising era of gene therapy for beta-hemoglobinopathies, hematological malignancy, and hemophilia. *Annals of Hematology*. 2024 Jun;103(6):1805-17.
 - 67 Camaschella C. Iron-deficiency anemia. *New England journal of medicine*. 2015 May 7;372(19):1832-43.
 - 68 Camaschella C. Iron-deficiency anemia. *New England journal of medicine*. 2015 May 7;372(19):1832-43.
 - 69 Kremyanskaya M, Ginzburg YZ, Hoffman R. Modulators of the Heparin Pathway in Polycythemia Vera and Myelofibrosis. *Blood Journal*. 2025 Oct 16:1028643.
 - 70 Mansour GK, Hajjar AW, Sajid MR. Therapeutic targeting of the hepcidin-ferroportin axis and erythropoietic modulators: a narrative review. *Frontiers in Medicine*. 2025 Dec 9;12:1726337.
 - 71 Nemeth E, Ganz T. Heparin-ferroportin interaction controls systemic iron homeostasis. *International journal of molecular sciences*. 2021 Jun 17;22(12):6493.
 - 72 Li L, Mandal PK. Recent advancements in gene therapy for sickle cell disease and β -thalassemia. *Frontiers in Hematology*. 2024 Sep 27;3:1468952.
 - 73 Alayoubi AM, Khawaji ZY, Mohammed MA, Mercier FE. CRISPR-Cas9 system: a novel and promising era of gene therapy for beta-hemoglobinopathies, hematological malignancy, and hemophilia. *Annals of Hematology*. 2024 Jun;103(6):1805-17.
 - 74 Sharma A, Kumar A, Saha PK, Saha L. The role of Tmprss6 gene polymorphism in iron resistance iron deficiency anaemia (IRIDA): a systematic review. *Annals of Hematology*. 2024 Apr;103(4):1085-102.

- 75 Williams DA, Kohn DB, Thrasher AJ. Ex vivo Modification of Hematopoietic Stem and Progenitor Cells for Gene Therapy. *Molecular Therapy*. 2025 Apr 1.
- 76 Locatelli F, Cavazzana M, Frangoul H, de la Fuente J, Algeri M, Meisel R. Autologous gene therapy for hemoglobinopathies: From bench to patient's bedside. *Molecular Therapy*. 2024 May 1;32(5):1202-18.
- 77 Alayoubi AM, Khawaji ZY, Mohammed MA, Mercier FE. CRISPR-Cas9 system: a novel and promising era of gene therapy for beta-hemoglobinopathies, hematological malignancy, and hemophilia. *Annals of Hematology*. 2024 Jun;103(6):1805-17.
- 78 Petzer V, Tymoszuk P, Asshoff M, Carvalho J, Papworth J, Deantonio C. A fully human anti-BMP6 antibody reduces the need for erythropoietin in rodent models of the anemia of chronic disease. *Blood, The Journal of the American Society of Hematology*. 2020 Aug 27;136(9):1080-90.
- 79 Nai A, Silvestri L, Asperti M, Vinchi F. Current landscape of hepcidin therapeutics. In *Iron Metabolism in Human Health and Disease* 2025 Jul 3 (pp. 399-418). Cham: Springer Nature Switzerland.
- 80 Ramos E, Ruchala P, Goodnough JB, Kautz L, Preza GC, Nemeth E. Minihepcidins prevent iron overload in a hepcidin-deficient mouse model of severe hemochromatosis. *Blood, The Journal of the American Society of Hematology*. 2012 Nov 1;120(18):3829-36.
- 81 Schwartz AJ, Goyert JW, Solanki S, Kerk SA, Chen B, Castillo C. Hepcidin sequesters iron to sustain nucleotide metabolism and mitochondrial function in colorectal cancer epithelial cells. *Nature Metabolism*. 2021 Jul;3(7):969-82.
- 82 Nemeth E, Ganz T. Hepcidin and iron in health and disease. *Annual review of medicine*. 2023 Jan 27;74(1):261-77.
